# Human kidney patient-derived organoids are a promising tool for modeling drug-induced nephrotoxicity and for assessing preclinical toxicity of immunotherapy



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#### Introduction

Toxicology studies are essential to assess drug safety and represent a pivotal step of the preclinical drug development. However, advances in drug discovery are often hampered by the lack of suitable preclinical models that recapitulate the in vivo physiology of the tissue. For instance, findings in animal models are not always translatable to humans due to species specificity, and in vitro cell lines might display abnormal expression of the relevant targets, enzymes, and transporters. Lastly, primary human cells from tissue explants have limited availability and are unstable in culture. Thus, there is a compelling need for advanced in vitro preclinical models to address drug-induced toxicity, especially for organs most frequently affected by toxicity, such as the kidneys. HUB Organoids<sup>®</sup> are innovative "mini-organs in a dish" derived from adult epithelial stem cells, which form 3D structures resembling the architecture of the epithelial tissue of origin and recapitulating the parental tissue physiology. They are genetically and phenotypically stable in culture and can be scaled up for screening purposes, providing a unique platform for toxicity studies. Here, we established kidney patient-derived organoids (PDOs) from the renal cortex, the portion of the kidney containing proximal tubules; these structures play a major role in eliminating waste products and can be particularly susceptible to drug-induced toxicity. For this reason, we employed our newly established models to assess the effect of known nephrotoxic compounds and to determine the safety profile of a T Cell Bispecific (TCB) antibody in a coculture assay with allogeneic T cells.

## TCB titration in cocultures of cortical PDOs and T cells



### **Methods**

- Human kidney Organoids establishment according to proprietary technology
- Nephrotoxin viability screens using ATP-based and live-imaging readouts
- T cell coculture assay using live-imaging and cytokine release quantification

#### Establishment of human kidney organoids

Human kidney organoids were successfully established from both the renal cortex and medulla from 6 different patients.

		Tissue	Organoids			
Kidney O	rganoid establishment	rtex	CORE CO	00	(P'G)	



 ESK1 TCB targeting the WT1 peptide presented by the HLA-A2 complex and CD3 led to increased organoid killing specifically in cocultures of HLA-A2 positive PDOs





# Nephrotoxin viability screens in cortical PDOs



- Increased IFN-γ (and Granzyme B) release specifically in cocultures of HLA-A2 positive PDOs
- No increased organoid killing, or cytokine release observed with a nontargeting (NT) TCB and in the cocultures of HLA-A2 negative PDOs.

## Summary

Kidney organoids:

- Can be successfully derived from different kidney regions
- Can be employed in high-throughput screening assays for assessing drug nephrotoxicity
- Can be cocultured with allogeneic T cells for determining immunotherapy safety

# Conclusions

HUB Organoids developed from patient-derived kidney tissue represent a promising model system for drug toxicity

HUB has developed a living biobank of kidney cortical PDOs that can be employed for large scale screening of nephrotoxic compounds as wells as immunotherapeutics, providing a valuable *in vitro* tool to predict drug-induced nephrotoxicity and to assess preclinical immunotoxicity.

#### References

- Consistent IC<sub>50</sub> values among different PDOs and experiments
  Analogous results from the ATP-based and Caspase3/7-based readouts
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